

Reliable interpretation from reliable data for complex proteomic characterization is a challenge

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LC-MSMS is a method of choice for protein identification in a complex mixture. However, the peak capacity of mono-dimensional LC is too low for very complex mixtures. Beside multidimensional LC, the use of exclusion lists combined with successive analyses has been shown to be very efficient to overcome this limitation. Even with high performance MS such as Orbitrap, the interpretation of the data is a challenge to sort out false positive results.

Material

3 samples : Proteins from the IF3 cell line (*Drosophila melanogaster*) Purified on agarose beads. 0,2 - Ing protein / sample

LC-MS/MS

Tryptic digestion : directly on the sepharose beads; peptides elution

Liquid chromatography on inverse phase column (Pepmap C18, 75 μm I.D., 15 cm lenght, Dionex) with a flow rate of 220 nL/min.

Material and Methods

Mass spectrometer hybrid linear ion trap /Orbitrap (LTQ Orbitrap, Thermofisher, San Jose, CA USA) with a nanospray ion source. Automatic acquisition between MS and MS/MS

Orbitrap resolution 6000, between 500 and 2000 Da Followed by 3 MS/MS scan (LTQ) on the 5 most intense peaks. Exclusion 90 sec of the fragmented precursors.

3 analysis / sample, followed by 3 analysis / sample with MS/MS exclusion of ≈200 peptids from the most abundant proteins (±10 ppm on whole LC analysis)

Results

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<u>Mascot conditions:</u> Score peptide >20, >30, >40 OR no filter. Search on Mudpit mode <u>Bioworks conditions:</u> Xc vs Charge : 1.5, 2.5, 3.0, 3.0 OR 1, 2, 3, 3 <u>For each sample :</u> 6 independant analysis and one <u>Merge</u> analysis (merging

6 independant analysis and one <u>Merge</u> analysis (merging the peptides from the 6 runs)

1/ Contribution of the kind of analysis



The use of an exclusion list allows the identification of proteins never found without exclusion. For the proteins identified by two peptides, the merge of the 6 runs allows the identification of new proteins, even with stringent criteria.

2/ Repetability



Repetability of the protein identification (3 runs without exclusion list). For each parameter used, the ratio of proteins found 3x vary between 1:3 and 1:2. In most of the cases, it is not possible to practic according to the protein score if the protein will be found 1x, 2x or 3x.



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In this example, the best « Bioworks peptide » was not found by Mascot.

Proteins w by both so proteins, s

Proteins with a very high score are found by both sofware, however, for the other proteins, some identifications are specifics of Bioworks and some other specific of Mascot.

4/ Effect of the search engine on peptides identification

3/ Effect of the search engine on protein identification



Y = 3.16x + 3.29 R² = 0.85

Mascat and Bioworks peptide score from a single run. A lot of low-score peptides are found only with one software. There is no simple rule to explain why some hight-score peptide are found only by one software. In black, linear regression for the peptide found by both software; in orange linear regression for all the peptides.



In this example, the best « Mascot peptide » was not identified by Bioworks because the software don't take in account the processing of the N-terminal M (it should be noted that this peptide is found if we are looking for partial trypsic digestion)

Conclusions

Our study underlines the positive effect of an exclusion list for the LC-MS/MS protein identification in a complex mixture.

The requirement of triplicate analysis and combined search engines for reliable identification has been suggested. The present work gives an estimation of the proteins excluded if we consider only the proteins found in triplicate or by both software.

At the peptide level, validation criteria for databases search parameters should be tuned very accurately so that different search engines will give similar results.

The validation criteria should be considered as a wholein an experimental design, according to the possibility -or not -to use an orthogonal method to validate the protein.



The ratio of modified peptides decreases with the peptide score in both software. This ratio is also correlated with the group of the peptide (found by both softwares or only one)